

CASE REPORT

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# Plasma and synovial fluid concentrations of linezolid in patients with knee osteoarthritis infected with *Staphylococcus aureus*

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## Abstract

**Background:** Linezolid is a new oxazolidinone antibiotic used for infections caused by methicillin-resistant *Staphylococcus* and other species.

**Case presentation:** Two cases of knee osteoarthritis with acute infection were successfully treated using linezolid. The plasma and synovial fluid concentrations of linezolid in two patients [women aged 69 and 73 years (cases 1 and 2)] with knee osteoarthritis infected with *Staphylococcus aureus* were measured after they were administered 600 mg twice daily by intravenous infusion. The plasma linezolid concentrations during knee surgery in case 1 at day 5 and in case 2 at day 2 were 19.6 and 15.6 µg/mL, respectively. The synovial fluid concentrations of linezolid in samples taken during surgery in case 1 and case 2 were 14.9 and 17.0 µg/mL, respectively; these values corresponded to ratios of synovial fluid/plasma of 76 and 109%. Possible metabolite 2-hydroxylated linezolid potentially mediated by cytochrome P450 2J2 was not detected in the plasma or synovial fluid samples under the current clinical setting after multiple doses.

**Conclusions:** These results implied nearly equivalent concentrations of linezolid in plasma and synovial fluid of clinical patients with knee osteoarthritis acutely infected with *Staphylococcus aureus*.

**Keywords:** Drug monitoring data, Knee osteoarthritis, Synovial fluid penetration

## Background

Linezolid is a new oxazolidinone antibiotic used for infections caused by methicillin-resistant *Staphylococcus* and other species [1, 2]. Linezolid is a promising antibiotic drug, but its use is limited by adverse effects with prolonged administration of 600 mg twice daily [3–5]. The pharmacokinetics of linezolid in healthy subjects [6] and in obese patients with cellulitis [7] have been reported. Linezolid is reportedly hydroxylated by as-yet

unidentified cytochrome P450s [8], but recently a role for P450 2J2 in linezolid 2-hydroxylation was indicated [9].

The case of a patient with chronic infection of an orthopedic implant treated with linezolid has been reported [10]. In the current study, two cases of acute infection of knee osteoarthritis were successfully treated using linezolid. From a clinical perspective, the monitoring of synovial fluid concentrations of linezolid is of interest. Synovial fluid concentrations of linezolid assessed during knee surgery in these two patients were determined and the ratios of synovial fluid/plasma were calculated.

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### Case presentation

Two cases of acute infection of knee osteoarthritis were successfully treated using linezolid between August 2018 and February 2019. The plasma and synovial fluid concentrations of linezolid in two patients (women aged 68 and 73 years, Table 1) treated in Kamakura Hospital for knee osteoarthritis infected with *Staphylococcus aureus* were determined after administration of 600 mg twice daily by intravenous infusion (at 6 am and 6 pm) for 11 days in case 1 and 19 days in case 2. Clinical laboratory results before linezolid treatment for case 1 were serum creatinine (0.61 mg/dL), aspartate aminotransferase (32 IU/L), alanine aminotransferase (37 IU/L), serum albumin (4.1 g/dL), and platelet ( $36.5 \times 10^4/\mu\text{L}$ ); and those for case 2 were serum creatinine (0.40 mg/dL), aspartate aminotransferase (22 IU/L), alanine aminotransferase (31 IU/L), serum albumin (3.0 g/dL), and platelet ( $30.0 \times 10^4/\mu\text{L}$ ). Co-administrated drugs during linezolid treatment for both cases were loxoprofen (180 mg per day from day 1) and lansoprazole (15 mg from day 1), with additional clindamycin (900 mg from day 2) for case 2. The patients were discharged 25 days (case 1) and 41 days (case 2) after admission. The patients gave written informed consent to take part in this study and for its publication. The Ethics Committee of Kamakura Hospital approved this study. The synovial fluid samples collected from the two patients during surgery along with plasma samples were pharmacokinetically analyzed. Samples (50  $\mu\text{L}$ ) of both plasma and synovial fluid taken from the two above-described patients were treated with an equal volume of acetonitrile, and the aqueous supernatant was centrifuged at  $15,000 \times g$  for 10 min at 4 °C.

Under the current clinical setting, it was focused to find any hydroxylated linezolid metabolite(s) in the plasma or synovial fluid samples. To investigate the possible P450 2J2-dependent 2-hydroxylation of linezolid [9], an incubation mixture was prepared consisting of 100 pmol/mL recombinant P450 2J2 protein (Corning, Woburn, MA, USA), 0.40 mM linezolid, an NADPH-generating system (0.50 mM NADP<sup>+</sup>, 5.0 mM glucose 6-phosphate, and 0.50 units/mL glucose 6-phosphate dehydrogenase), and 100 mM potassium phosphate buffer (pH 7.4) in a

total volume of 0.20 mL. The reaction was carried out at 37 °C for 60 min and was terminated by adding 0.60 mL of acetonitrile. The reaction mixture was then centrifuged at  $15,000 \times g$  for 10 min.

Plasma and synovial fluid concentrations of linezolid and that of the in vitro product of linezolid and P450 2J2 as a possible metabolite were determined using a reverse-phase high-performance liquid chromatography system (Shimadzu, Kyoto, Japan) with a Mightysil RP-18GP Aqua column (5  $\mu\text{m}$ ,  $150 \times 4.6 \text{ mm}$ , Kanto Chemical, Tokyo, Japan) equilibrated in a mobile phase comprising 10% CH<sub>3</sub>CN in 0.1% aqueous formic acid at a flow rate of 1.5 mL/min and a column temperature of 45 °C with ultraviolet monitoring at 254 nm. This mobile phase composition was held for 1 min followed by a linear gradient to 50% CH<sub>3</sub>CN at 8 min, a second linear gradient to 62% CH<sub>3</sub>CN at 10 min, a 5 min wash at 90% CH<sub>3</sub>CN, followed by re-equilibration at the initial conditions for another 4 min. Samples (10  $\mu\text{L}$ ) were infused using an autosampler. The retention times of linezolid and its possible 2-hydroxylated metabolite [9] were 7.4 and 5.9 min respectively. Data are given as means and standard deviations from triplicate determinations.

The clinical laboratory results for these two patients over a 2-week period are shown in Fig. 1A and B. The patient in case 2 was found by outsourced clinical laboratory services to be suffering from methicillin-resistant *Staphylococcus aureus* infection, but in case 1, the infectious agent was methicillin-sensitive *Staphylococcus aureus*. For cases 1 and 2, the measured plasma concentrations of linezolid and the pharmacokinetic-modeled concentration profiles of linezolid (obtained using a previously reported one-compartment model with the following parameters of absorption constant,  $0.126 \text{ h}^{-1}$ ; volume of distribution, 22 L; and half-life, 5.5 h) are shown in Fig. 1C and D, respectively [11]. The plasma linezolid concentrations in samples taken during knee surgery in case 1 at day 5 and in case 2 at day 2 were 19.6 and 15.6  $\mu\text{g/mL}$ , respectively.

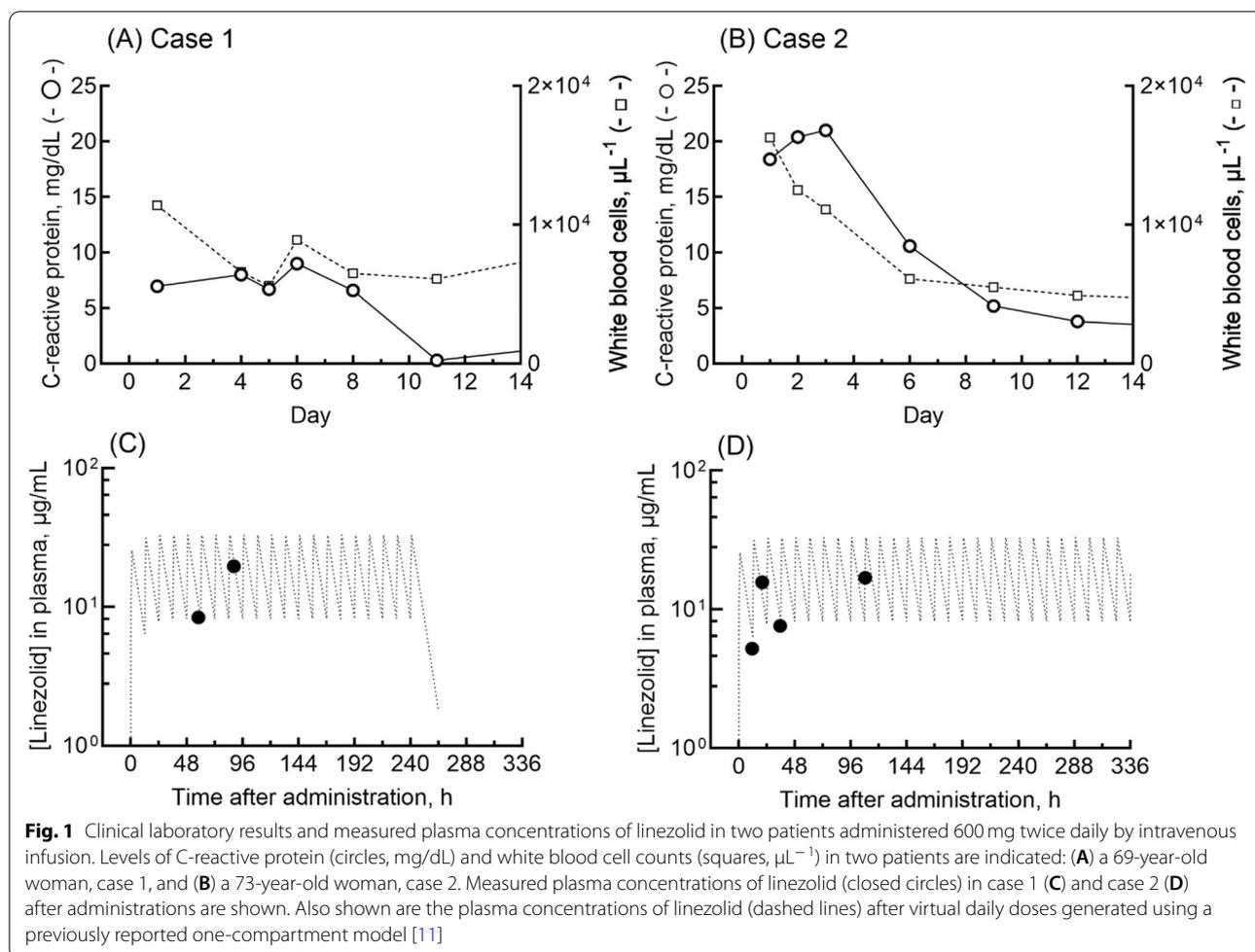
Figure 2 shows representative chromatograms for linezolid in plasma and synovial fluid. There were additional polar peaks to that of substrate linezolid alone

**Table 1** Plasma and synovial fluid concentrations of linezolid determined in two patients during knee surgery

| Case (body weight)          | Sampling | Linezolid concentration ( $\mu\text{g/mL}$ ) |                | Ratio of synovial fluid/plasma |
|-----------------------------|----------|----------------------------------------------|----------------|--------------------------------|
|                             |          | Plasma                                       | Synovial fluid |                                |
| 1 69-year-old woman (50 kg) | Day 5    | $19.6 \pm 0.9$                               | $14.9 \pm 0.4$ | 76%                            |
| 2 73-year-old woman (50 kg) | Day 2    | $15.6 \pm 0.2$                               | $17.0 \pm 0.5$ | 109%                           |

Data are means  $\pm$  standard deviations from triplicate determinations

The linezolid levels in plasma and synovial fluid samples from the two patients (Fig. 1) were quantified using liquid chromatography

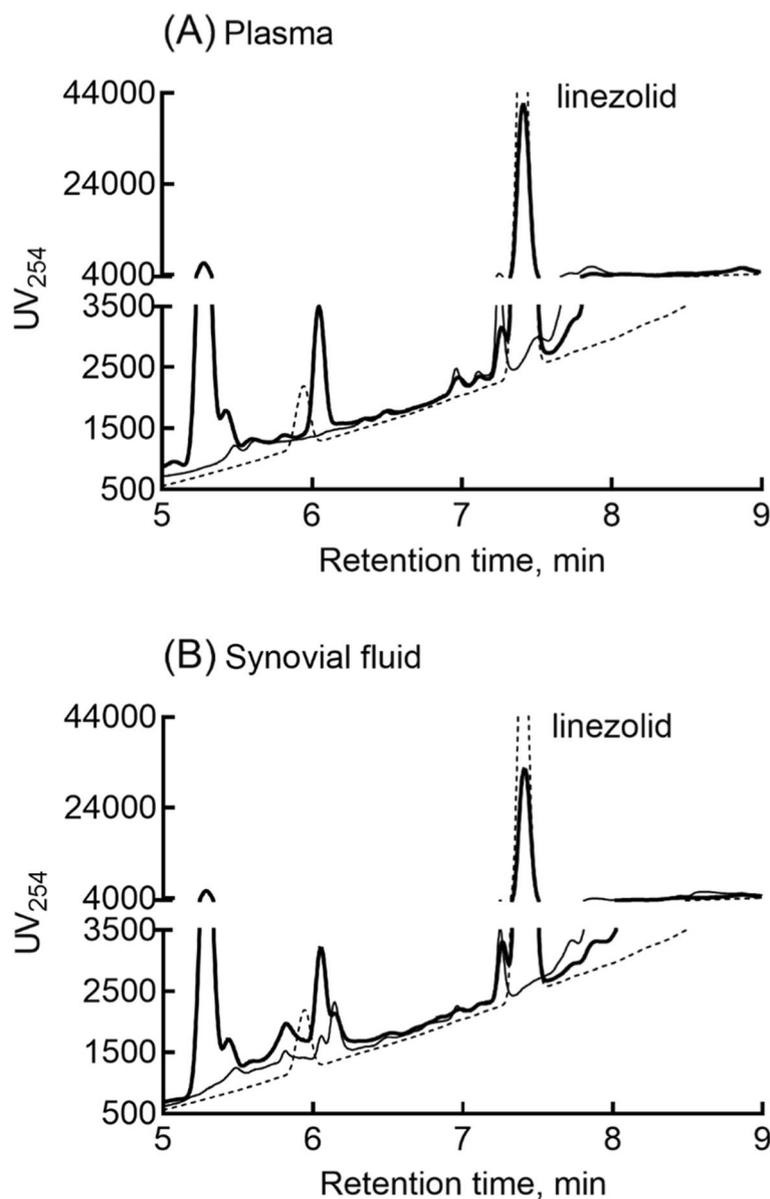


in both plasma and synovial fluid samples from the patient after infusion of linezolid. However, the presence of 2-hydroxylated linezolid potentially mediated by cytochrome P450 2J2 was not detected in these plasma or synovial fluid samples in the current clinical setting (Fig. 2). Synovial fluid concentrations of linezolid taken during knee surgery in case 1 and case 2 were 14.9 and 17.0  $\mu\text{g}/\text{mL}$ , respectively, which corresponded to ratios of synovial fluid/plasma of 76 and 109% (Table 1).

### Discussion and conclusion

Linezolid is reportedly not highly protein-bound (30%) and has a moderate volume of distribution similar to the total water content of the body [12]. Relatively good penetration of linezolid into inflammatory blister fluid has been demonstrated in healthy volunteers [13, 14]. It also has been reported that linezolid concentrations in soft tissue specimens ranged from 18 to 78% (mean, 51%) of their simultaneous serum levels in diabetic patients with foot infections [11]. Plasma-to-synovial-fluid ratios of  $0.76 \pm 0.34$  for linezolid have been

reported in terms of the knee gap after a single dose of 600 mg of linezolid [15]. Although the package insert information of linezolid suggests the presence of two minor morpholine ring-opening metabolites in human plasma or urine (in-house document), recently reported 2-hydroxylated linezolid (mediated by cytochrome P450 2J2 [9]) was not considered so far, to our knowledge, in the fluid samples in the clinical setting. In the current study, similar plasma and synovial fluid concentrations of parent compound linezolid in two patients with knee osteoarthritis infected with *Staphylococcus aureus* were demonstrated after patients were administered multiple doses of 600 mg by twice daily intravenous infusion. The present results suggested almost equivalent concentrations of linezolid in plasma and synovial fluid of clinical patients with knee osteoarthritis infected with *Staphylococcus aureus* after multiple doses. A population-based pharmacokinetic model of linezolid in hospitalized patients with chronic arthritis has been proposed [16]. Simple systematic therapeutic drug monitoring for linezolid [17], which may reflect



**Fig. 2** Representative chromatograms for linezolid in plasma and linezolid in synovial fluid. Plasma (A) and synovial fluid (B) samples from case 1 before (thin lines) and after (thick lines) treatment with linezolid twice a day were investigated. Possible linezolid 2-hydroxylated metabolite(s) mediated by recombinant human P450 2J2 in an in vitro system (dashed lines) was used for chromatographic comparison. The retention times of linezolid and its possible 2-hydroxylated metabolite [9] were 7.4 and 5.9 min respectively

the concentrations of linezolid in synovial fluid after multiple doses, would have a beneficial clinical impact on the treatment of synovial fluid infections in patients with knee osteoarthritis.

#### Abbreviation

P450: cytochrome P450.

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#### Authors' contributions

D. Negishi, O. Mitsumatsu, T. Matsumura, H. Mitsumatsu monitored the patients and carried out the acquisition of patient data. D. Negishi, M. Makiguchi, and M. Shimizu mainly carried out the pharmacokinetic analyses.

D. Negishi and H. Yamazaki designed the study and mainly wrote the manuscript. All authors gave final approval of the manuscript. The author(s) read and approved the final manuscript.

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#### Availability of data and materials

All data generated or analyzed during this study are included in this published article and are also available from the corresponding author on reasonable request.

#### Declarations

##### Ethics approval and consent to participate

This study was approved by the Ethics Committee of Kamakura Hospital. Informed consent was obtained from the patient.

##### Consent for publication

Not Applicable.

##### Competing interests

The authors declare that they have no competing interests.

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#### References

- Hashemian SMR, Farhadi T, Ganjparvar M. Linezolid: a review of its properties, function, and use in critical care. *Drug Des Devel Ther*. 2018;12:1759–67.
- Steinmetz MP, Vogelbaum MA, De Georgia MA, Andrefsky JC, Isada C. Successful treatment of vancomycin-resistant enterococcus meningitis with linezolid: case report and review of the literature. *Crit Care Med*. 2001;29:2383–5.
- Krull M, Klare I, Ross B, Trenchel R, Beelen DW, Todt D, et al. Emergence of linezolid- and vancomycin-resistant enterococcus faecium in a department for hematologic stem cell transplantation. *Antimicrob Resist Infect Control*. 2016;5:31.
- Stein GE, Wells EM. The importance of tissue penetration in achieving successful antimicrobial treatment of nosocomial pneumonia and complicated skin and soft-tissue infections caused by methicillin-resistant *Staphylococcus aureus*: vancomycin and linezolid. *Curr Med Res Opin*. 2010;26:571–88.
- Baccani I, Antonelli A, Galano A, Bartalesi F, Bartoloni A, Rossolini GM. Linezolid-resistant enterococcus faecalis infection following prolonged low-dosage linezolid treatment for multidrug-resistant tuberculosis. *Clin Infect Dis*. 2017;65:2159–60.
- Slatter JG, Stalker DJ, Feenstra KL, Welshman IR, Bruss JB, Sams JP, et al. Pharmacokinetics, metabolism, and excretion of linezolid following an oral dose of [(14)C] linezolid to healthy human subjects. *Drug Metab Dispos*. 2001;29:1136–45.
- Stein GE, Schooley SL, Peloquin CA, Kak V, Havlicek DH, Citron DM, et al. Pharmacokinetics and pharmacodynamics of linezolid in obese patients with cellulitis. *Ann Pharmacother*. 2005;39:427–32.
- Wynalda MA, Hauer MJ, Wienkers LC. Oxidation of the novel oxazolidinone antibiotic linezolid in human liver microsomes. *Drug Metab Dispos*. 2000;28:1014–7.
- Obach RS. Linezolid metabolism is catalyzed by cytochrome P450 2J2, 4F2, and 1B1. *Drug Metab Dispos*. 2022;50:413–21.
- Dell'Aquila AM, Janovsky C, Cohen M. Case report - infection of Total knee arthroplasty treated with one-stage surgery and linezolid. *J Bone Jt Infect*. 2017;2:163–6.
- Stein GE, Schooley S, Peloquin CA, Missavage A, Havlicek DH. Linezolid tissue penetration and serum activity against strains of methicillin-resistant *Staphylococcus aureus* with reduced vancomycin susceptibility in diabetic patients with foot infections. *J Antimicrob Chemother*. 2007;60:819–23.
- MacGowan AP. Pharmacokinetic and pharmacodynamic profile of linezolid in healthy volunteers and patients with gram-positive infections. *J Antimicrob Chemother*. 2003;51(Suppl 2):ii17–25.
- Dehghanyar P, Burger C, Zeitlinger M, Islinger F, Kovar F, Muller M, et al. Penetration of linezolid into soft tissues of healthy volunteers after single and multiple doses. *Antimicrob Agents Chemother*. 2005;49:2367–71.
- Gee T, Ellis R, Marshall G, Andrews J, Ashby J, Wise R. Pharmacokinetics and tissue penetration of linezolid following multiple oral doses. *Antimicrob Agents Chemother*. 2001;45:1843–6.
- Schwameis R, Syre S, Sarahrudi K, Appelt A, Marhofer D, Burau D, et al. Penetration of linezolid into synovial fluid and muscle tissue after elective arthroscopy. *J Antimicrob Chemother*. 2017;72:2817–22.
- Tsuji Y, Holford NHG, Kasai H, Ogami C, Heo YA, Higashi Y, et al. Population pharmacokinetics and pharmacodynamics of linezolid-induced thrombocytopenia in hospitalized patients. *Br J Clin Pharmacol*. 2017;83:1758–72.
- Galar A, Valerio M, Munoz P, Alcalá L, García-González X, Burillo A, et al. Systematic therapeutic drug monitoring for linezolid: variability and clinical impact. *Antimicrob Agents Chemother*. 2017;61:e00687–17.

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