CASE REPORT Open Access

Plasma infliximab monitoring contributes to optimize Takayasu arteritis treatment: a case report



Sho Masui^{1,2}, Atsushi Yonezawa^{1,2*}, Kazushi Izawa³, Makoto Hayakari¹, Kayoko Asakura¹, Risa Taniguchi¹, Masahiko Isa³, Hirofumi Shibata³, Takahiro Yasumi³, Ryuta Nishikomori³, Junko Takita³ and Kazuo Matsubara¹

Abstract

Background: Infliximab (IFX), a mouse-human chimeric monoclonal antibody against human tumor necrosis factor alpha, is used in refractory cases of Takayasu arteritis. Several factors influence the pharmacokinetics of therapeutic antibodies including IFX. Monitoring plasma levels of IFX could be a useful approach in optimizing treatment via individual dose adjustment.

Case presentation: Here, we report the case of a 4-year-old Takayasu arteritis girl who was resistant to standard therapy. IFX was started at 5 mg/kg (day 0). C-reactive protein (CRP) levels decreased from 8.7 (day 0) to 1.6 mg/dL (day 10). CRP levels were thereafter elevated again on day 23 (9.0 mg/dL), and body fluid leakage at the inflammation site in the legs was observed. Trough IFX levels decreased from 23.6 (day 10) to 2.5 µg/mL (day 23). Based on the trough levels, IFX was given biweekly at 8 mg/kg. Plasma IFX levels gradually increased, and CRP levels decreased to around 2 mg/dL. A similar pattern -initial decreases followed by increases- was observed between clinical course of IFX and IgG levels. It was speculated that IgG and IFX losses were due to fluid leakage from the patient's necrotizing legs.

Conclusions: Monitoring of plasma IFX levels can be a potential tool to optimize the treatment in Takayasu arteritis patients.

Keywords: Infliximab, Pharmacokinetics, Takayasu arteritis, LC-MS/MS

Background

Infliximab (IFX), a mouse-human chimeric monoclonal antibody against human tumor necrosis factor alpha (TNF- α), is used in the treatment of several autoimmune diseases. It has been reported that several factors influence the pharmacokinetics of therapeutic antibodies, such as development of anti-drug antibodies (ADAs) [1–3] and nephropathy [4]. Monitoring plasma IFX levels could be a potential tool for optimizing treatment via individual dose adjustment [5–7]. In fact, a tool (RemicheckQ $^{\circ}$) with a similar purpose has been already approved for measuring blood concentrations of IFX. RemicheckQ $^{\circ}$ is a diagnostic kit used to determine whether serum IFX concentration is

less or more than $1\,\mu g/mL$ in patients with rheumatoid arthritis in Japan. However, monitoring of IFX levels is not common in other diseases. Takayasu arteritis is an autoimmune nonspecific large vasculitis affecting the aorta and its main branches with unknown etiology. Based on the Guidelines for Management of Vasculitis Syndrome [8] and reports [9–11], anti-TNF- α agents (such as IFX) are also used in refractory cases of Takayasu arteritis. Here, we report the case of a 4-year-old girl with Takayasu arteritis, in whom monitoring of plasma IFX levels was useful as a means of adjusting the regimen of IFX administration.

¹Department of Clinical Pharmacology and Therapeutics, Kyoto University Hospital, 54 Shogoin Kawahara-cho, Sakyo-ku, Kyoto 606-8507, Japan ²Graduate School of Pharmaceutical Sciences, Kyoto University, 54 Shogoin Kawahara-cho, Sakyo-ku, Kyoto 606-8507, Japan Full list of author information is available at the end of the article



Case presentation

A 4-year-old Japanese girl had fever and swelling in the right leg, with marked elevation of C-reactive protein (CRP) levels. Based on computed tomography, echocardiography and skin biopsy, she had been diagnosed with

^{*} Correspondence: ayone@kuhp.kyoto-u.ac.jp

Takayasu arteritis at the age of two years. Due to aggravated inflammation, blood flow decreased in her legs, and part of her right leg became necrotic. As she had been resistant to standard therapy with prednisolone or tocilizumab without monitoring plasma concentrations, we started to administer IFX (day 0). IFX was given at a dose of 5 mg/kg on days 0 and 10. Although the levels initially decreased from 8.7 (day 0) to 1.6 mg/dL (day 10), CRP contents elevated again on day 23 (9.0 mg/dL), and IFX was administered at 10 mg/kg on the same day. Body fluid leakage from the inflammation sites in her legs was observed. Because blood IgG levels were lower than standard value, immunoglobulin (2.5 g) has been administered on days 17, 31, 37, 45, 51, 59, 65, 72, 85 and thereafter once a week for at least a few months.

Plasma IFX concentrations were measured by LC-MS/MS with nano-surface and molecular-orientation limited (nSMOL, Shimadzu, Kyoto, Japan) proteolysis [12, 13]. Based on the clinical courses of blood CRP and IFX levels (Fig. 1), trough IFX levels were decreased from 23.6 μ g/mL (day 10) to 2.5 μ g/mL (day 23). Dosages and intervals of IFX administrations were then adjusted

according to the trough IFX levels. IFX was given biweekly at 8 mg/kg per administration. Plasma IFX levels gradually increased, and CRP levels decreased to around 2 mg/dL 40 days after IFX administration. Inflammation was suppressed, and the dosage of prednisolone could be gradually decreased. CRP levels transiently elevated to 5.8 and 7.0 mg/dL after infection on days 87 and 126, respectively. Blood culture results confirmed the presence of gram-positive cocci. ADA against IFX was not detected using the enzyme-linked immunosorbent assay kit (Somru BioScience, PEI, Canada). During the observation period, no renal failure was observed. It is noteworthy to mention that a similar pattern -initial decreases followed by increases- was observed in the clinical courses of IFX and IgG (Fig. 1).

Discussion

The present report described treatment of a young female Takayasu arteritis patient with IFX, resistant to standard therapy of tocilizumab without monitoring plasma concentrations. Because IFX therapy is an off-label use for the treatment of Takayasu arteritis, the

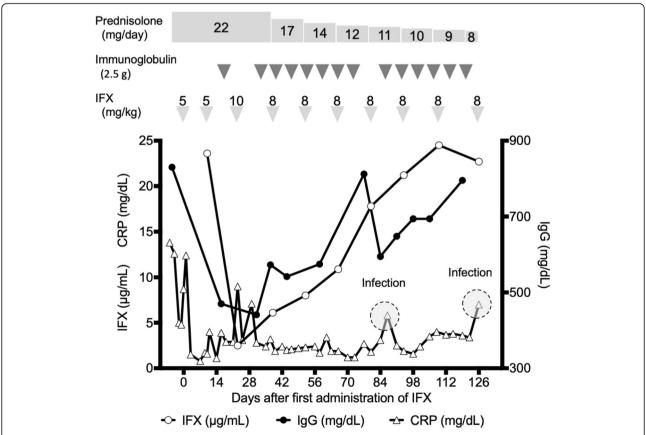


Fig. 1 Clinical course of the patient. Trough plasma infliximab (IFX, open circle), immunoglobulin G (IgG, closed circle) and C-reactive protein levels (CRP, open triangle) levels are shown. IFX was administered at 5 mg/kg (days 0 and 10), 10 mg/kg (day 23), and thereafter at 8 mg/kg at 2-week intervals. Immunoglobulin (2.5 g) was administered on days 17, 31, 37, 45, 51, 59, 65, 72, 85 and thereafter once a week for at least a few months. Gray circles represent the escalation of CRP on days 87 and 126 due to infections

treatment regimen was based on that for inflammatory bowel disease: i.e. 5 mg/kg at weeks 0, 2, 6 and subsequently at 8-week intervals. Wolbink et al. [14] have reported that the median (interquartile range) trough IFX levels in rheumatoid arthritis patients (dose: 3 mg/kg) at weeks 2, 6, 14 registered 22.3 (15.3–29.4), 14.6 (7.3–22) and 2.8 (0.6-6.8) µg/mL, respectively. However, in our case, the trough IFX levels were 2.5 µg/mL on day 23 (3-4 weeks) and $6.1 \mu\text{g/mL}$ on day 38 (5-6 weeks), even though IFX was administered at 5 mg/kg on days 0 and 10, and 10 mg/kg on day 23. It was suggested that IFX levels were too low to suppress the inflammation during this period. Finally, IFX administration at 8 mg/kg per 2 weeks succeeded in maintaining sufficient IFX levels to suppress the inflammation. This is the first report showing the relation to plasma IFX concentrations and effects on Takayasu arteritis. Monitoring of IFX levels was useful in the treatment of this patient with Takayasu arteritis.

Several factors affect pharmacokinetics of therapeutic antibodies, such as development of ADA and nephropathy. Many reports have indicated that consequent ADA formation due to an immunogenicity of IFX may decrease the functional drug concentration, resulting in a loss of response [1-3]. Counsilman et al. [4] have also reported that rituximab is rapidly excreted in the urine of a patient with severe nephrosis. However, ADA against IFX was not detected in the present patient, and no renal failure was also observed. Interestingly, there was an analogous tendency between the clinical courses of IFX and IgG levels. When both IFX and IgG levels decreased at around day 23, the patient had severe inflammation in her legs with substantial exudate. Although there were no previous reports, it was speculated that there was a loss of IgG (including IFX) due to the leakage from the necrotizing sites. Unfortunately, we could not collect the body fluid or measure IFX or IgG in it. Thereafter, a fluid leakage gradually decreased in association with wound healing, and the concentrations of both IFX and IgG elevated at the same dosage. This suggests that blood IgG level can be used as an index for monitoring IFX concentration.

Conclusions

Inadequate IFX administration caused therapeutic failure. Monitoring of plasma IFX levels can be a useful approach in optimizing treatment of Takayasu arteritis patients.

Abbreviations

ADA: anti-drug antibody; CRP: C-reactive protein; IFX: Infliximab; IgG: Immunoglobulin G; LC-MS/MS: Liquid chromatography tandem mass spectrometry; nSMOL: nano-surface and molecular-orientation limited; TNF- α : Tumor necrosis factor alpha

Acknowledgments

The authors thank Dr. T. Shimada and Dr. N. Iwamoto from Leading Technology of Bioanalysis and Protein Chemistry, SHIMADZU Corporation for some technical supports and discussion in the experiments with nSMOL and LC-MS/MS.

Funding

This work was supported by the Research on Regulatory Harmonization and Evaluation of Pharmaceuticals, Medical Devices, Regenerative and Cellular Therapy Products, Gene Therapy Products, and Cosmetics from the Japan Agency for Medical Research and Development (AMED), the Japan Research Foundation for Clinical Pharmacology, and the Mochida Memorial Foundation for Medical and Pharmaceutical Research to A.Y.

Availability of data and materials

Data used in this case report will not be shared due to the risk of identifying an individual.

Authors' contributions

AY an KI conceived the study and designed the protocol. SM, AY, KI, MH and KM contributed to writing the manuscript. KI, KA, RT, MI, HS, TY, RN and JT monitored the patients and carried out acquisition of the patient data. SM measured the plasma concentration of IFX. As a follow-up, SM, AY and MH analyzed and interpreted the data. All authors reviewed the final manuscript.

Ethics approval and consent to participate

All procedures performed in studies involving human participants were executed in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards. This study was approved by the Ethics Committee of Kyoto University Graduate School and Faculty of Medicine and Kyoto University Hospital (R1386). Written informed consent was obtained from the patient's mother for this case report.

Consent for publication

A copy of the written consent is available for review by the Editor-in-Chief of this journal.

Competing interests

The authors declare that they have no competing interest.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Author details

¹Department of Clinical Pharmacology and Therapeutics, Kyoto University Hospital, 54 Shogoin Kawahara-cho, Sakyo-ku, Kyoto 606-8507, Japan. ²Graduate School of Pharmaceutical Sciences, Kyoto University, 54 Shogoin Kawahara-cho, Sakyo-ku, Kyoto 606-8507, Japan. ³Department of Pediatrics, Graduate School of Medicine, Kyoto University, 54 Shogoin Kawahara-cho, Sakyo-ku, Kyoto 606-8507, Japan.

Received: 18 January 2019 Accepted: 4 April 2019 Published online: 02 May 2019

References

- Baert F, Noman M, Vermeire S, Van Assche G, G DH, Carbonez A, Rutgeerts
 P. Influence of immunogenicity on the long-term efficacy of infliximab in
 Crohn's disease. N Engl J Med. 2003;348:601–8.
- Pecoraro V, De Santis E, Melegari A, Trenti T. The impact of immunogenicity of TNFalpha inhibitors in autoimmune inflammatory disease. A systematic review and meta-analysis. Autoimmun Rev. 2017;16:564–75.
- Strand V, Balsa A, Al-Saleh J, Barile-Fabris L, Horiuchi T, Takeuchi T, Lula S, Hawes C, Kola B, Marshall L. Immunogenicity of biologics in chronic inflammatory diseases: a systematic review. BioDrugs. 2017;31:299–316.
- Counsilman CE, Jol-van der Zijde CM, Stevens J, Cransberg K, Bredius RG, Sukhai RN. Pharmacokinetics of rituximab in a pediatric patient with therapy-resistant nephrotic syndrome. Pediatr Nephrol. 2015;30:1367–70.
- Takeuchi T, Miyasaka N, Inoue K, Abe T, Koike T. Impact of trough serum level on radiographic and clinical response to infliximab plus methotrexate

- in patients with rheumatoid arthritis: results from the RISING study. Mod Rheumatol. 2009;19:478–87.
- Price S. Rheumatoid arthritis: monitoring serum concentration of infliximab might improve RA disease control. Nat Rev Rheumatol. 2010;6:66.
- Vaughn BP, Martinez-Vazquez M, Patwardhan VR, Moss AC, Sandborn WJ, Cheifetz AS. Proactive therapeutic concentration monitoring of infliximab may improve outcomes for patients with inflammatory bowel disease: results from a pilot observational study. Inflamm Bowel Dis. 2014;20:1996–2003.
- Guideline for management of vasculitis syndrome (JCS 2008). Japanese Circulation Society. Circ J 2011, 75:474–503.
- Hoffman GS, Merkel PA, Brasington RD, Lenschow DJ, Liang P. Anti-tumor necrosis factor therapy in patients with difficult to treat Takayasu arteritis. Arthritis Rheum. 2004;50:2296–304.
- Molloy ES, Langford CA, Clark TM, Gota CE, Hoffman GS. Anti-tumour necrosis factor therapy in patients with refractory Takayasu arteritis: longterm follow-up. Ann Rheum Dis. 2008;67:1567–9.
- Comarmond C, Plaisier E, Dahan K, Mirault T, Emmerich J, Amoura Z, Cacoub P, Saadoun D. Anti TNF-alpha in refractory Takayasu's arteritis: cases series and review of the literature. Autoimmun Rev. 2012;11:678–84.
- Iwamoto N, Shimada T, Umino Y, Aoki C, Aoki Y, Sato TA, Hamada A, Nakagama H. Selective detection of complementarity-determining regions of monoclonal antibody by limiting protease access to the substrate: nanosurface and molecular-orientation limited proteolysis. Analyst. 2014;139:576–80.
- Iwamoto N, Yokoyama K, Takanashi M, Yonezawa A, Matsubara K, Shimada T. Verification of LC-MS bioanalysis by nSMOL in human serum between original and biosimilar therapeutic antibody infliximab. Curr Pharm Biotechnol. 2018.
- Wolbink GJ, Voskuyl AE, Lems WF, de Groot E, Nurmohamed MT, Tak PP, Dijkmans BA, Aarden L. Relationship between serum trough infliximab levels, pretreatment C reactive protein levels, and clinical response to infliximab treatment in patients with rheumatoid arthritis. Ann Rheum Dis. 2005;64:704–7.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

